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NO:9), J (SEQ ID NO:10), K (SEQ ID NO:11), L (SEQ ID NO:12), M (SEQ ID NO:13). --

Please replace the Figure description starting at page 5, line 5 with the following rewritten Figure description:

- Figures 2A and 2B

Oligonucleotide sequences and analogs thereof complementary to bcl- X_L mRNA; Figure 2A shows: A (SEQ ID NO:14), A'(SEQ ID NO:15), B (SEQ ID NO:16), C (SEQ ID NO:17), C'(SEQ ID NO:18), D (SEQ ID NO:19), E (SEQ ID NO:20), E'(SEQ ID 21), F (SEQ ID NO:22), G (SEQ ID NO:23), G'(SEQ ID NO:24); Figure 2B shows, H (SEQ ID NO:25), H'(SEQ ID NO:26), I (SEQ ID NO:27), I'(SEQ ID NO:28), J (SEQ ID NO:29), K (SEQ ID NO:30), K'(SEQ ID NO:31), L (SEQ ID NO:32), L'(SEQ ID NO:33), M (SEQ ID NO:34), M'(SEQ ID NO:35).

Please replace the Figure description starting at page 6, line 2 with the following rewritten Figure description:

--Figure 10

Most active chimeric PS-PO oligonucleotides by their ability to down-regulate Bcl-XL protein expression (from top to bottom SEQ ID NO:15; SEQ ID NO:19; SEQ ID NO:24; SEQ ID NO:27; SEQ ID NO:22; SEQ ID NO:29; and SEQ ID NO:38).

In the Specification:

Please replace the paragraph starting page 22, line 33 with the following rewritten paragraph:

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- Two clones of LNCaP cells overexpressing bcl-xL protein (1072-4 and 1072-5) have been obtained after transfection of wild type LNCaP cells with the plasmid vector pSFFV/bcl-xL and lipofectin. Also a mock transfectant clone of LNCaP cells carrying $\text{neo}^{\text{@}}$ resistance gene (1072-3) was used for the control experiments. Clone 1072-4 demonstrates 10-fold overexpression, and clone 1072-5 - 4-fold overexpression of bcl-xL protein. Western blot analysis for bcl-xL protein was performed as described above. Results for bcl-xL protein expression were confirmed by Northern blot analysis for bcl-xL mRNA expression, demonstrating significant elevation of this mRNA in bcl-xL transformed cell lines. For the Northern blot analysis, the total RNA was isolated from the cells using TRIZOL reagent (GIBCO BRL), and 20 $\mu\mathrm{g}$ aliquotes were separated RNA-formaldehyde gel, blotted onto nylon membranes (Schleicher & Schull), UV-linked and prehybridized for two hours at 42 °C in the standard hybridization solution. Then the blot was hybridized overnight with the PCR-amplified fragment of human bcl-xL cDNA at 42 °C. Bcl-xL coding fragment was amplified from pSFFV/bcl-xL plasmid using bcl-x specific primers. The primer sequences were: bcl-x-upstream, 5'-ATGTCTCAGAGCAACCGGGA-3' (SEQ ID NO:36): bcl-x-downstream, 5'-TCATTTCCGACTGAAGAGTG-3'(SEQ ID NO:37). Twenty five cycles of amplification were performed in DNA Thermal Cycler (Perkin-Elmer, Norwalk, CT) at 94 °C (30 sec), $^{\circ}\text{C}$ (30 sec), and 72 $^{\circ}\text{C}$ (30 sec). The PCR products were analysed on a 1.2% agarose gel. The resultant fragment was labeled by random primer method to the specific activity 10^7 cpm/ng of the probe and used for the hybridization. After washings blots were autoradiographed for 24h at -80 $^{\circ}\text{C.}$ Blots

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